

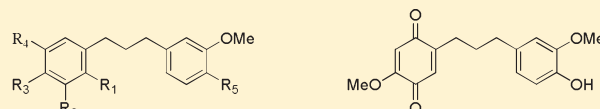
Diarylpropanes and an Arylpropyl Quinone from *Combretum griffithii*

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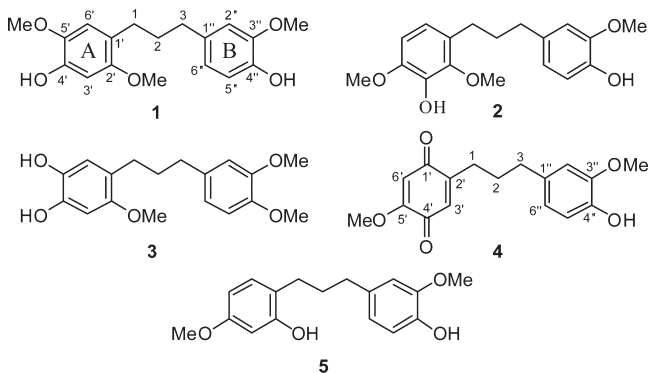
S Supporting Information

ABSTRACT: Three new diarylpropanes (**1–3**), a new arylpropyl quinone (**4**), and the known 1-(2-hydroxy-4-methoxyphenyl)-3-(4-hydroxy-3-methoxyphenyl)propane (**5**) were isolated from a methanol extract of stems of *Combretum griffithii*. Their structures were elucidated by spectroscopic methods. Compounds **1**, **2**, **4**, and **5** showed cytotoxicity against one or more cancer cell lines (KB, MCF7, and NCI-H187), and compound **5** exhibited activity against *Mycobacterium tuberculosis* (MIC 3.13 $\mu\text{g}/\text{mL}$).



- 1;** R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OMe, R₅ = OH
2; R₁ = OMe, R₂ = OH, R₃ = OMe, R₄ = H, R₅ = OH
3; R₁ = OMe, R₂ = H, R₃ = OH, R₄ = OH, R₅ = OMe
5; R₁ = OH, R₂ = H, R₃ = OMe, R₄ = H, R₅ = OH

Combretum griffithii Van Heurck & Müll. Arg. (Combretaceae) is a vine, known as “Khamin khrua”¹ in Thai, and a water decoction of the stem has been used traditionally by local people as a treatment for hepatitis.² No previous phytochemical study of *C. griffithii* has been reported. However, previous investigations of *Combretum* species resulted in the isolation of triterpenes, triterpene glucosides,^{3,4} pentacyclic triterpenes,^{5,6} triterpenoids with 9 β ,19-cyclopropyl-1-en-3-one skeletons,⁷ cyclobutane dimers,⁸ cycloartane triterpenes,^{9,10} and ellagic acid derivatives.¹¹ As part of our work on bioactive constituents of Thai plants, a MeOH extract of stems of *C. griffithii* showed cytotoxicity against the KB oral human epidermal carcinoma cell line (IC₅₀ 2.6 $\mu\text{g}/\text{mL}$). We report herein the isolation, structural elucidation, and bioactivity of three new diarylpropanes (**1–3**) and a new arylpropyl quinone (**4**), named griffithanes A–D, together with the known diarylpropane **5** from *C. griffithii*. It should be noted that **5** was the first diarylpropane reported from the family Combretaceae.



RESULTS AND DISCUSSION

Separation of a MeOH extract of *C. griffithii* by silica gel column chromatography (CC) and preparative TLC yielded four new compounds (**1–4**) and 1-(2-hydroxy-4-methoxyphenyl)-3-(4-hydroxy-3-methoxyphenyl)propane (**5**).¹² Compound **1**

was a viscous liquid, and its molecular formula was deduced as C₁₈H₂₂O₅ from HRESITOFMS (observed m/z 319.1548 [M + H]⁺), indicating eight degrees of unsaturation. The IR spectrum showed absorption bands for OH (3419 cm⁻¹) and aromatic (3000, 1612, and 1512 cm⁻¹) groups. The ¹H and ¹³C NMR spectra and a DEPT experiment showed 18 carbon signals attributable to three methoxy, three methylene, five methine aromatic, and seven nonprotonated carbons. The ¹H NMR spectrum of **1** showed a –CH₂–CH₂–CH₂– unit at δ 2.56 (t, J = 8.0 Hz, H-1), 1.89 (quint, J = 8.0 Hz, H-2), and 2.60 (t, J = 8.0 Hz, H-3), which was confirmed by the COSY spectrum. The two benzene rings showed resonances indicating a 1,2,4,5-tetrasubstituted ring A [δ 6.63 (s, H-6') and 6.41 (s, H-3')] and a 1,3,4-trisubstituted ring B [δ 6.69 (br s, H-2''), 6.83 (d, J = 8.4 Hz, H-5''), and 6.68 (br d, J = 8.4 Hz, H-6'')]. Three OCH₃ signals were also observed [δ 3.77 (s), 3.80 (s), and 3.85 (s)]. The ¹³C NMR spectrum exhibited five resonances at low field [δ _C 147.4, 147.8, 142.9, 146.4, and 143.6] indicating oxygenated ring carbons. The HMBC spectrum demonstrated correlations of H-3' to C-2', C-4', and C-5'; H-6' to C-5', C-4', C-2', and C-1; methoxy protons at δ _H 3.77 to C-2'; and methoxy protons at δ _H 3.80 to C-5', consistent with a 1,2,4,5-tetrasubstituted ring A. HMBC correlations of H-2'' to C-4'' and C-6''; H-5'' to C-1'', C-3'', and C-4''; H-6'' to C-2'' and C-4''; H-3 to C-1'', C-2'', and C-6''; and methoxy protons at δ _H 3.85 to C-3'' confirmed the connection of ring B. The NOESY spectrum exhibited correlations between H-3' and methoxy protons at C-2'; H-6' and methoxy protons at C-5'; and H-2'' and methoxy protons at C-3''. On the basis of the above data, compound **1** was defined as a new compound, 1-(4-hydroxy-2,5-dimethoxyphenyl)-3-(4-hydroxy-3-methoxyphenyl)propane, which was named griffithane A.

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Compound **2** was assigned the molecular formula $C_{18}H_{22}O_5$, as deduced from HRESITOFMS data (observed m/z 319.1547 $[M + H]^+$). The IR spectrum indicated the presence of OH (3425 cm^{-1}) and aromatic (3000 , 1605 , and 1511 cm^{-1}) groups. The ^1H and ^{13}C NMR and DEPT spectra of **2** were similar to those of **1**, except that the aromatic protons of ring A showed doublets at δ 6.64 ($J = 8.4\text{ Hz}$, H-5') and 6.77 ($J = 8.4\text{ Hz}$, H-6'), indicating that ring A of **2** was a 1,2,3,4-tetrasubstituted benzene ring. The HMBC spectrum showed correlations indicating OCH_3 groups at C-2' and C-4' and an OH group at C-3'. Thus, compound **2** was a new compound, 1-(3-hydroxy-2,4-dimethoxyphenyl)-3-(4-hydroxy-3-methoxyphenyl)propane, and it was named griffithane B.

Compound **3** had the molecular formula $C_{18}H_{22}O_5$ as deduced from HRESITOFMS. Absorption bands in the IR spectrum at 3424 , 1605 , and 1506 cm^{-1} indicated the presence of OH and aromatic groups. The ^1H and ^{13}C NMR spectra, and a DEPT experiment, revealed that **3** was a 1,3-diarylpropane containing three OCH_3 groups that were similar to those of **1** and **2**, except for the different positions on both benzene rings. An OCH_3 group at C-2' and OH groups at C-4' and C-5' on aromatic ring A were deduced from the HMBC correlations of H-3' to C-1', C-2', and C-4', and $\text{CH}_3\text{O}-2'$ to C-2', as well as from NOESY correlations between $\text{CH}_3\text{O}-2'$ and H-3' and between OH protons at C-4' and C-5'. In addition, the ^{13}C NMR resonances showed downfield shifts of C-2' (δ_{C} 150.9), C-4' (δ_{C} 144.7), and C-5' (δ_{C} 139.2), while C-3' appeared upfield (δ_{C} 97.4) due to electronic effects from the *ortho* position of methoxy and hydroxy groups. Benzene ring B contained two OCH_3 groups at C-3'' (δ_{C} 146.3) and C-4'' (δ_{C} 143.5), which were confirmed by the HMBC correlations of $\text{CH}_3\text{O}-3''$ to C-3'' and $\text{CH}_3\text{O}-4''$ to C-4''. Thus, **3** was defined as 1-(4,5-dihydroxy-2-methoxyphenyl)-3-(3,4-dimethoxyphenyl)propane, which was named griffithane C.

Compound **4** was obtained as orange crystals and was assigned the molecular formula $C_{17}H_{18}O_5$ from HRESITOFMS (observed m/z 325.1047 $[M + \text{Na}]^+$), indicating nine degrees of unsaturation. The IR data showed the presence of OH (3450 cm^{-1}) and conjugated dicarbonyl (1673 and 1647 cm^{-1}) groups. The ^{13}C NMR and DEPT data exhibited 17 carbons attributable to two methoxy, three methylene, five methine, and seven nonprotonated (including two carbonyl ketone) carbons. The ^1H and ^{13}C NMR spectra revealed a 1,3-disubstituted propane having an aromatic ring B similar to that of **1**; however, ring A was different. The ^1H NMR spectrum of **4** showed signals corresponding to two olefinic protons at δ_{H} 6.45 (s, H-3') and 5.87 (s, H-6'). The ^{13}C NMR spectrum showed the presence of two carbonyl carbons [δ_{C} 182.2 and 187.4] and four olefinic carbons [δ_{C} 150.3 (C-2'), 130.5 (C-3'), 158.5 (C-5'), and 107.7 (C-6')], which indicated a benzoquinone unit. The HMBC spectrum showed correlations of H-3' to C-1 and C-5'; H-6' to C-1', C-2', C-4', and C-5'; and methoxy protons to C-5'. NOESY correlations of H-3' with H-1 and methoxy protons at C-5' with H-6' confirmed the 1,4-benzoquinone moiety containing an OCH_3 group at C-5' and connected to the propyl chain at C-1. Thus, **4** was elucidated as a new compound, 1-[2-(5-methoxy-1,4-benzoquinone)]-3-(4-hydroxy-3-methoxyphenyl)propane, which was named griffithane D.

Compounds **1**, **2**, **4**, and **5** exhibited cytotoxicity against the KB cancer cell line with IC_{50} values of 2.13, 5.67, 1.42, and 2.18 $\mu\text{g}/\text{mL}$, respectively. Compound **4** also showed cytotoxic activity against NCI-H187 and MCF7 cancer cell lines, with IC_{50} values of 1.08 and 6.75 $\mu\text{g}/\text{mL}$, respectively. These assays indicate that

the OCH_3 group at C-3'' and OH group at C-4'' (*ortho*-substituted on ring B in **1**, **2**, **4**, and **5**) contribute to the cytotoxic activity against the KB cancer cell line. The benzoquinone of ring A might also play an important role in enhancing cytotoxicity. In addition, compound **5** displayed antimycobacterial activity against *Mycobacterium tuberculosis*, with a MIC of 3.13 $\mu\text{g}/\text{mL}$. Compounds **1**–**5** were tested for antiplasmodial (antimalarial) activity in vitro against *Plasmodium falciparum*, but they were all inactive at 10 $\mu\text{g}/\text{mL}$.

EXPERIMENTAL SECTION

General Experimental Procedures. Melting points were determined using a Gallenkamp melting point apparatus and are uncorrected. UV spectra were measured on an Agilent 8453 UV–visible spectrophotometer. IR spectra were recorded on a Bruker Tensor 27 FT-IR spectrophotometer. NMR spectra were recorded in CDCl_3 on a Varian Mercury Plus 400 spectrometer, using residual CHCl_3 as an internal reference. HRESITOFMS were obtained using a Micromass Q-TOF-Z spectrometer. Column chromatography and preparative TLC were carried out on silica gel 60 (230–400 mesh) and PF_{254} , respectively.

Plant Material. Stems of *C. griffithii* were collected from Khon Kaen Province, Thailand, in June 2009, and were identified by James F. Maxwell, Department of Biology, Chiang Mai University. A voucher specimen (SRITUBTIM 61) was deposited at the Udon Thani Rajabhat University Herbarium, Thailand.

Extraction and Isolation. Air-dried stems of *C. griffithii* (2.7 kg) were ground and extracted successively at room temperature with hexane (3 L \times 3), EtOAc (3 L \times 3), and MeOH (3 L \times 3) to give crude hexane (5.3 g), EtOAc (20.3 g), and MeOH (136.7 g) extracts.

The MeOH extract (61.8 g) was separated initially by silica gel CC, eluted with a gradient system of hexane–EtOAc and EtOAc–MeOH. Each fraction (100 mL) was monitored by TLC, and fractions with similar TLC patterns were combined to yield six fractions (F_1 – F_6). Fraction F_1 was subjected to silica gel flash CC, eluted with a gradient of CH_2Cl_2 –MeOH to furnish seven subfractions ($F_{1/1}$ – $F_{1/7}$). Subfraction $F_{1/2}$ was separated by preparative TLC (EtOAc–hexane, 1:3, three times) to yield **1** (10.0 mg), **2** (79.7 mg), and **4** (46.8 mg). Subfraction $F_{1/3}$ was purified by preparative TLC (EtOAc–hexane, 1:1.5, three times) to afford **5** (79.0 mg) and an additional amount of **2** (41.6 mg). Preparative TLC of subfraction $F_{1/4}$ (EtOAc–hexane, 1:1.5, three times) gave an additional amount of **5** (11.0 mg). Preparative TLC of $F_{1/7}$ (EtOAc–hexane, 1:1.5, three times) afforded **3** (20.0 mg). Fraction F_2 was subjected to flash CC, eluted with MeOH– CH_2Cl_2 (1:50), and further separated by preparative TLC (EtOAc–hexane, 1:1.5) to yield an additional amount of **1** (8.3 mg).

Griffithane A (1): viscous liquid; UV (MeOH) λ_{max} ($\log \epsilon$) 204 (4.63), 288 (3.69) nm; IR (neat) ν_{max} 3419, 3000, 2935, 2856, 1612, 1512, 1450, 1413, 1268, 1198, 1151, 1108, 1032, 998, 931, 853, 819 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 6.83 (1H, br d, $J = 8.4\text{ Hz}$, H-5''), 6.69 (1H, br s, H-2''), 6.68 (1H, d, $J = 8.4\text{ Hz}$, H-6''), 6.63 (1H, s, H-6'), 6.41 (1H, s, H-3'), 3.85 (3H, s, OCH_3-3''), 3.80 (3H, s, OCH_3-5'), 3.77 (3H, s, OCH_3-2'), 2.60 (2H, t, $J = 8.0\text{ Hz}$, H-3), 2.56 (2H, t, $J = 8.0\text{ Hz}$, H-1), 1.89 (2H, quint, $J = 8.0\text{ Hz}$, H-2); ^{13}C NMR (100 MHz, CDCl_3) δ 147.8 (C-4'), 147.4 (C-2'), 146.4 (C-3''), 143.6 (C-4''), 142.9 (C-5'), 134.2 (C-1''), 120.9 (C-6''), 119.0 (C-1'), 114.2 (C-5'), 113.9 (C-6'), 111.0 (C-2''), 101.0 (C-3'), 56.7 (OCH_3-5'), 55.9 (OCH_3-2'), 55.8 (OCH_3-3''), 35.5 (C-3), 31.8 (C-2), 29.0 (C-1); HRESITOFMS m/z 319.1548 $[M + H]^+$ (calcd for $C_{18}H_{22}O_5 + H$, 319.1545).

Griffithane B (2): viscous liquid; UV (MeOH) λ_{max} ($\log \epsilon$) 204 (5.11), 288 (4.15) nm; IR (neat) ν_{max} 3425, 3000, 2934, 2842, 1605, 1511, 1451, 1267, 1231, 1199, 1119, 1032, 856, 818 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 6.82 (1H, d, $J = 8.4\text{ Hz}$, H-5''), 6.77 (1H, d, $J = 8.4\text{ Hz}$, H-6'), 6.68 (2H, m, H-2'' and H-6''), 6.64 (1H, d, $J = 8.4\text{ Hz}$,

H-5'), 3.91 (3H, s, OCH₃-4'), 3.87 (3H, s, OCH₃-3''), 3.82 (3H, s, OCH₃-2'), 2.59 (2H, t, J = 8.0 Hz, H-3), 2.56 (2H, t, J = 8.0 Hz, H-1), 1.86 (2H, quint, J = 8.0 Hz, H-2); ¹³C NMR (100 MHz, CDCl₃) δ 150.7 (C-2'), 147.6 (C-3'), 146.2 (C-3''), 143.5 (C-4''), 139.7 (C-4'), 134.3 (C-1''), 127.5 (C-1'), 124.4 (C-6'), 120.9 (C-6''), 114.1 (C-5''), 111.0 (C-2''), 110.0 (C-5'), 60.6 (OCH₃-4'), 60.4 (OCH₃-2'), 55.8 (OCH₃-3''), 35.3 (C-3), 32.7 (C-2), 29.1 (C-1); HRESITOFMS *m/z* 319.1547 [M + H]⁺ (calcd for C₁₈H₂₂O₅ + H, 319.1545).

Griffithane C (3): viscous liquid; UV (MeOH) λ_{max} (log ε) 204 (4.68), 288 (3.72) nm; IR (neat) ν_{max} 3424, 3001, 2934, 2842, 1605, 1506, 1451, 1371, 1265, 1118, 1030, 853, 816 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.82 (1H, d, J = 8.0 Hz, H-5''), 6.73 (2H, s, H-6' and H-2''), 6.68 (1H, d, J = 8.0 Hz, H-6''), 6.48 (1H, s, H-3'), 5.45 (1H, s, OH), 5.17 (1H, s, OH), 3.88 (3H, s, OCH₃-3''), 3.87 (3H, s, OCH₃-4''), 3.77 (3H, s, OCH₃-2'), 2.57 (2H, t, J = 8.0 Hz, H-3), 2.55 (2H, t, J = 8.0 Hz, H-1), 1.84 (2H, quint, J = 8.0 Hz, H-2); ¹³C NMR (100 MHz, CDCl₃) δ 150.9 (C-2'), 146.3 (C-3''), 144.7 (C-4'), 143.5 (C-4''), 139.2 (C-5'), 134.6 (C-1''), 123.4 (C-1'), 120.9 (C-6''), 115.9 (C-6'), 114.5 (C-5''), 111.1 (C-2''), 97.4 (C-3'), 56.5 (OCH₃-2'), 55.8 (OCH₃-3''), 55.2 (OCH₃-4''), 35.2 (C-3), 31.9 (C-2), 29.0 (C-1); HRESITOFMS *m/z* 319.1544 [M + H]⁺ (calcd for C₁₈H₂₂O₅ + H, 319.1545).

Griffithane D (4): orange crystals (EtOAc); mp 121–123 °C; UV (MeOH) λ_{max} (log ε) 203 (3.99), 266 (3.63) nm; IR (neat) ν_{max} 3450, 3065, 3017, 2930, 2848, 1673, 1647, 1599, 1513, 1454, 1364, 1269, 1122, 1031, 957, 851, 818 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.78 (1H, d, J = 8.0 Hz, H-5''), 6.65 (1H, d, J = 8.0 Hz, H-6''), 6.64 (1H, s, H-2''), 6.45 (1H, s, H-3'), 5.87 (1H, s, H-6'), 3.84 (3H, s, OCH₃-3''), 3.77 (3H, s, OCH₃-5'), 2.58 (2H, t, J = 7.6 Hz, H-3), 2.43 (2H, t, J = 7.6 Hz, H-1), 1.79 (2H, quint, J = 7.6 Hz, H-2); ¹³C NMR (100 MHz, CDCl₃) δ 187.4 (C-1'), 182.2 (C-4'), 158.5 (C-5'), 150.3 (C-2'), 146.5 (C-3''), 143.8 (C-4''), 133.2 (C-1''), 130.5 (C-3'), 120.9 (C-6''), 114.3 (C-5''), 110.9 (C-2''), 107.7 (C-6'), 56.2 (OCH₃-5'), 55.9 (OCH₃-3''), 35.1 (C-3), 29.8 (C-2), 28.6 (C-1); HRESITOFMS *m/z* 325.1047 [M + Na]⁺ (calcd for C₁₇H₁₈O₅ + Na, 325.1052).

Antimalarial Assay. Antimalarial activity was evaluated in vitro against the parasite *Plasmodium falciparum* (K1, multi-drug-resistant strain), using the method of Trager and Jensen.¹³ Quantitative assessment of malarial activity in vitro was determined by means of the microculture radioisotope technique based upon the method described by Desjardins et al.¹⁴ The inhibitory concentration (IC₅₀) represents the concentration that causes 50% reduction in parasite growth as indicated by the in vitro uptake of [³H]-hypoxanthine by *P. falciparum*. The standard compound dihydroartemisinin exhibited an IC₅₀ value of 1.0 ng/mL.

Antimycobacterial Assay. Antimycobacterial activity was assessed against *Mycobacterium tuberculosis* H37Ra using the microplate Alamar Blue assay (MABA).¹⁵ The standard drug isoniazid showed MIC values of 0.23–0.46 μg/mL.

Cytotoxicity Assay. Cytotoxicity assays against human epidermoid carcinoma (KB), human breast cancer (MCF7), and human small cell lung cancer (NCI-H187) cell lines were performed employing the colorimetric method as described by Skehan and co-workers.¹⁶ The reference substance was doxorubicine.

■ ASSOCIATED CONTENT

Supporting Information. ¹H and ¹³C NMR spectra for 1–4 are available free of charge via the Internet at <http://pubs.acs.org>.

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